# CHROMANS FROM HELIANTHELLA QUINQUENERVIS

### WERNER HERZ and PALANIAPPAN KULANTHAIVEL

Department of Chemistry, The Florida State University, Tallahassee, FL 32306, U.S.A.

(Received 9 May 1983)

Key Word Index—Helianthella quinquenervis; Compositae; Heliantheae; p-hydroxyacetophenone derivatives; encecalin; 3,4-chromanediols.

Abstract—Investigation of Helianthella quinquenervis afforded, in addition to several known 6-acetylchromenes, 3-prenyl-4-hydroxyphenone and the benzofuran encecalin, cis- and trans-2,2-dimethyl-3,4,7-trihydroxy-6-acetylchroman.

### INTRODUCTION

Helianthella is a small genus of helianthoid composites indigenous to the western United States and adjacent Canada and Mexico [1]. Transitionally it is included in subtribe Verbesininae (or Helianthinae) as defined by Stuessy [2], while a recent revision of the tribe has placed it in a redefined subtribe Ecliptinae [3]. Chemical reports on the genus seem to be limited to the roots of H. uniflora (Nutt.) T. & G. [4]. The present report describes the results of our examination of the aerial parts of H. quinquenervis (Hook.) Gray.

## RESULTS AND DISCUSSION

Chloroform extraction of H. quinquenervis furnished the known chromenes 1a-1c and the related known compounds 2 and 3. Substances 1a, 1b, 2 and 3 were previously isolated from the roots of H. uniflora [4] and 1c (encecalin) from the aerial parts of Encelia californica [5]. Two new isomeric diols 4a and 5a,  $C_{13}H_{16}O_5$ , were obtained in small amounts from the polar fraction. The <sup>1</sup>H NMR spectrum of 4a exhibited signals of mutually coupled protons at  $\delta 4.77$  and 3.73, shifted to  $\delta 6.59$  and 5.77 on conversion of 4a to dibenzoate 4b and hence obviously due to two hydrogens under vicinal hydroxyl groups. This, in conjunction with the other signals in the <sup>1</sup>H NMR spectrum (see Experimental), led to formulation of the diol as 4a, the cis-stereochemistry of the hydroxyl groups being based on the value of  $J_{3,4}$  (4 Hz). This was confirmed by partial synthesis of 4a by oxidation of 1a with osmium tetroxide.

The isomeric diol exhibited mutually coupled signals at  $\delta 4.50$  and 3.53 (J = 9 Hz) which moved downfield to  $\delta 6.44$  and 5.75 on dibenzoylation to **5b**; hence it was the trans-isomer **5a**. Attempts to synthesize **5a** from **1a** by the Prévost reaction gave the trans-iodoacetates **6a** and **6b**, but no diacetate. The facile iodination of the aromatic ring under these conditions is surprising; that C-5 was unsubstituted was shown by the chemical shift of the

Kis and coworkers [6] have isolated similar diols, the 3,4-cis- and 3,4-trans-dihydroxy-6-methoxy-2,2-dimethylchromans 8 and 9, from the basidomycete Panus rudis Fr. The absolute configurations shown in formulae 8 and 9 were established by CD spectrometry of the corresponding dibenzoates. Our diols were optically inactive.\*

From a chemical point of view, the root extract of H. uniflora and the extract of the herb of H. quinquenervis do not differ significantly. Further work is necessary to establish whether the occurrence of isopentenylacetophenones is characteristic of the genus. Some other representatives of genera are included in Ecliptinae as redefined [3] elaborate similar compounds, but the chemistry of the subtribe as a whole is not uniform [7].

### **EXPERIMENTAL**

Isolation of H. quinquenervis constituents. Above-ground parts (1.3 kg) of Helianthella quinquenervis (Hook.) Gray, collected by Dr. B. H. Braun in Boulder County, Colorado in summer 1960, were extracted with CHCl<sub>3</sub> and worked up in the usual fashion [8]. The crude gum (22 g) was adsorbed on 30 g silicic acid (Mallinckrodt 100 mesh) and chromatographed over 400 g of the same adsorbent packed in hexane, 500 ml fractions of eluant being collected as follows: fractions 1-4 (hexane), 5-10 (hexane-EtOAc, 19:1), 11-16 (hexane-EtOAc, 9:1), 17-22 (hexane-EtOAc, 4:1), 23-28 (hexane-EtOAc, 3:2), 29-32 (hexane-EtOAc, 2:3), 33-36 (hexane-EtOAc, 1:4), 37-38 (EtOAc), 39-40 (EtOAc-MeOH, 99:1) 41-42 (EtOAc-MeOH, 49:1), 43-44 (EtOAc-MeOH, 19:1) and 45-46 (EtOAc-MeOH, 9:1).

Fractions 5 and 6 were combined; recrystallization gave 3.4 g of 1a, mp  $81-82^\circ$ , lit. mp  $77^\circ$  [4]. Purification of fractions 5 and 6 by TLC ( $C_6H_6$ -EtOAc 19:1, multiple developments) afforded gummy 1b (0.355 g) and 1c (0.557 g). Fraction 12 had mainly one constituent; purification by TLC ( $C_6H_6$ -EtOAc, 19:1) yielded 2 (1.3 g); fraction 11 also gave 0.230 g 2. Purification of fraction 17 by TLC ( $C_6H_6$ -EtOAc 19:1, 2 developments) gave 3 (89 mg), mp  $89-91^\circ$  (Et<sub>2</sub>O-hexane), lit. mp  $93-94^\circ$  [4]. Fraction 25 on standing in EtOAc deposited 84 mg of a 1:1 mixture of 4a and 5a.

remaining aromatic proton signal which is allylically coupled to H-4. Prévost reaction on 1d afforded only 6c; base treatment of the latter resulted in the formation of 7 rather than hydrolysis to 5a, thus establishing the location of the non-aromatic iodine atom in 6a-6c.

<sup>\*</sup>The slight rotation exhibited by 5a (see Experimental) can be ascribed to an impurity as the rotation of 5b, like that of 4b, was 0° and as the CD curves of 4b and 5b were transparent.

Purification of 45 g of the mixture by TLC (CHCl<sub>3</sub>-MeOH-EtOAc, 18:1:1, 2 developments) yielded spectroscopically pure 4a (14 mg) and 5a (18 mg).

Diol 4a, mp 220–221° (MeOH) had IR bands (KBr) at 3470, 3400, 1650 and 1610 cm<sup>-1</sup>; <sup>1</sup>H NMR spectrum (270 MHz, CDCl<sub>3</sub> and 3 drops of DMSO- $d_6$ –D<sub>2</sub>O):  $\delta$ 7.97 (d, J = 1 Hz, H-5), 6.34 (H-8), 4.77 (dd, J = 4, 1 Hz, H-4), 3.73 (d, J = 4 Hz, H-3), 2.58 (3H, H-9), 1.50 and 1.31 (br, each 3H, H-10 and H-11), and 12.42 (7-OH). [Calc. for C<sub>13</sub>H<sub>16</sub>O<sub>5</sub>: MW, 252.0999. Found: MW(MS), 252.1013.]

Diol 5a had mp 170–172° (CHCl<sub>3</sub>–MeOH),  $[\alpha]_D$  – 6°, IR bands (KBr) at 3470, 3400, 1650 and 1610 cm<sup>-1</sup>; <sup>1</sup>H NMR spectra (270 MHz, CDCl<sub>3</sub>–3 drops of DMSO- $d_6$ ):  $\delta$ 12.41 (7-OH), 7.96 (d, J = 1 Hz, H-5), 6.25 (H-8), 4.91 (d, J = 5 Hz, 4-OH), 4.82 (d, J = 4 Hz, 3-OH), 4.49 (ddd, J = 9, 5, 1 Hz, H-4), 3.52 (dd, J = 9, 4 Hz, H-3), 2.58 (3H, H-9), 1.49 and 1.24 (br, each 3H, H-10 and H-11) (addition of D<sub>2</sub>O caused disappearance of the –OH signals and simplification of the H-3 and H-4 signals); [Calc. for C<sub>13</sub>H<sub>16</sub>O<sub>5</sub>: MW, 252.0999. Found: MW(MS), 252.0986].

Benzoylation of the diols. The 1:1 mixture of 4a and 5a (30 mg) in 0.5 ml pyridine was stirred with 0.2 ml benzoyl chloride overnight at room temp. The usual work-up and purification of the product by TLC ( $C_6H_6$ -EtOAc, 39:1) afforded 31 mg 4b and 26 mg 5b. Dibenzoate 4b had mp 168-170° (Et<sub>2</sub>O-hexane),  $[\alpha]_D$  0°; <sup>1</sup>H NMR spectrum (270 MHz, CDCl<sub>3</sub>):  $\delta$ 8.26-7.30 (m, 16H, aromatic H and H-5), 6.84 (H-8), 6.59 (dd, J=4, 1 Hz, H-4), 5.77 (d, J=4 Hz, H-3), 2.45 (3H, H-9) and 1.57 (br, 6H, H-10 and H-11). Dibenzoate 5b had mp 191-191.5° (Et<sub>2</sub>O-hexane),  $[\alpha]_D$  0°; <sup>1</sup>H NMR spectrum (270 MHz, CDCl<sub>3</sub>):  $\delta$ 8.27-7.74 and 7.76-7.38 (m, 16H, aromatic H and H-5), 6.83 (H-8), 6.44 (dbr, J=58 Hz, H-4), 5.67 (d, J=58 Hz, H-3), 2.44 (3H, H-9), 1.61 and 1.54 (br, each 3H, H-10 and H-11).

Reactions of 1a. (A) A soln of 0.250 g 1a in 3 ml pyridine was stirred with 0.250 g O<sub>5</sub>O<sub>4</sub> at room temp. for 24 hr. After addition of 0.450 g NaHSO<sub>3</sub> in 8 ml H<sub>2</sub>O and 5 ml pyridine, stirring was continued for 15 min. The mixture was extracted with CHCl<sub>3</sub>. The washed and dried organic layer on evapn gave a product identical in all respects with 4a.

- (B) A soln of 0.400 g 1a in 15 ml  $C_6H_6$  was stirred with 0.80 g AgOAc in  $N_2$  while 0.60 g  $I_2$  was added in portions during a 10 min period. The mixture was refluxed for 2 hr, filtered while hot and evapd. TLC ( $C_6H_6$ -EtOAc, 39:1, 3 developments) of the residue yielded 0.120 g 6a and 0.210 g of a mixture of 6a and 6b. The low resolution MS of 6a had peaks at m/z (rel. int.) 530 ( $M^+$ ,  $C_{15}H_{16}O_5I_2$ ), 488 (2), 470 (19), 455 (46), 360 (73), 343 (100) and 328 (93);  $^1H$  NMR spectrum (270 MHz, CDCl<sub>3</sub>):  $\delta$ 13.54 (7-OH), 7.61 (br, H-5), 6.33 (dbr, J=7.5 Hz, H-4), 4.42 (d, J=7.5 Hz, H-3), 2.56 (3H, H-9), 2.22 (3H, Ac), 1.76 and 1.63 (br, each 3H, H-10 and H-11);  $^1H$  NMR spectrum of 6b (from that of the mixture, 270 MHz, CDCl<sub>3</sub>):  $\delta$ 12.44 (7-OH), 7.59 (br, H-5), 6.38 (H-8), 6.29 (dbr, J=7.5 Hz, H-4), 4.43 (d, J=7.5 Hz, H-3), 2.52 (3H, H-9), 2.21 (3H, Ac), 1.67 and 1.57 (each 3H, H-10 and H-11).
- (C) Acetylation of 1a furnished monoacetate 1d [4]. Repetition of the AgOAc–I<sub>2</sub> reaction with 1d (0.340 g) gave 0.240 g 6c;  $^1$ H NMR spectrum (270 MHz, CDCl<sub>3</sub>):  $\delta$ 7.70 (br, H-5), 6.60 (H-8), 6.35 (dbr, J=7.5 Hz, H-4), 4.24 (d, J=7.5 Hz, H-3), 2.48 (3H, H-9), 2.34 and 2.21 (each 3H, Ac), 1.68 and 1.59 (br, each 3H, H-10 and H-11). A soln of 60 mg 6c in 3.5 ml THF and 6 ml MeOH was stirred with 2 ml 1 N NaOH for 3 hr at room temp. Removal of the solvent gave 34 mg 7,  $^1$ H NMR spectrum

(270 MHz, CDCl<sub>3</sub>):  $\delta$ 12.73 (7-OH), 7.27 (H-5), 6.95 (H-4), 6.35 (H-8), 2.53 (3H, H-9) and 1.59 (6H, H-10 and H-11).

Acknowledgement—This work was supported in part by a grant (CA-13121) from the U.S. Public Health Service through the National Cancer Institute.

### REFERENCES

- 1. Weber, W. A. (1952) Am. Mid. Nat. 48, 1.
- Stuessy, T. F. (1977) in The Biology and Chemistry of the Compositae (Heywood, V. H., Harborne, J. B. and Turner, B. L., eds.), p. 621. Academic Press, London.
- 3. Robinson, H. (1981) Smithsonian Contrib. Bot. No. 51.
- 4. Bohlmann, F. and Grenz, M. (1970) Chem. Ber. 103, 90.
- Bjeldanes, L. and Geissman, T. A. (1969) Phytochemistry 8, 1293.
- Kis, Z., Closse, A., Sigg, H. P., Hruban, L. and Snatzke, G. (1970) Helv. Chim. Acta 53, 1577.
- Bohlmann, F., Tsankova, E., Jakupovic, J., King, R. M. and Robinson, H. (1983) Phytochemistry 22, 557.
- 8. Herz, W. and Högenauer, G. (1962) J. Org. Chem. 27, 1905.